



REVIEW ARTICLE

Modes of Action of Taurine and Granulocyte Colony-stimulating Factor in Neuroprotection

Chandana Buddhala, Howard Prentice, Jang-Yen Wu*

Department of Biomedical Science, Charles E. Schmidt College of Medicine, Florida Atlantic University, Boca Raton, Florida 33431, USA

ARTICLE INFO

Article history:

Received: Sep 22, 2011

Accepted: Oct 19, 2011

KEY WORDS:

apoptosis;
G-CSF;
G-CSF receptors;
neuroprotection;
taurine;
taurine receptors

New therapeutic targets are becoming increasingly popular for the treatment of a wide array of neurodegenerative diseases, the preferred targets being those that prevent neuronal apoptosis at multiple levels or those that can cross the blood-brain barrier in order to replace degenerated cells and promote neuronal regeneration. One such rapidly emerging neuroprotective agents is taurine. Taurine is a ubiquitous amino acid that satisfies most criteria to be classified as a neurotransmitter. Because of a wide spectrum of effects that taurine can induce on intrinsic apoptosis pathways, such as modulating mitochondrial pore permeability, attenuating endoplasmic reticulum stress, maintaining calcium homeostasis, and downregulating the activities of a range of pro-apoptotic proteins, including calpain and caspases, while upregulating a variety of anti-apoptotic proteins involved in glutamate and hypoxia-induced toxicity, taurine is being extensively studied and successfully applied for the treatment of neurodegenerative diseases. Another potential molecule being researched for combating neurodegenerative diseases is granulocyte colony-stimulating factor (G-CSF), which originates from the cytokine family of growth factors. G-CSF has gained widespread attention because of its ability to cross the blood-brain barrier, the presence of its receptors in the central nervous system, anti-apoptotic functions, and its proliferative role in the restoration of tissue survival via neurogenesis. In this review from the available current literature, the modes of action of taurine and G-CSF are discussed. Further mechanistic studies are warranted in order to fully realize the potential of these two molecules.

Copyright © 2011, Taipei Medical University. Published by Elsevier Taiwan LLC. All rights reserved.

1. Introduction

Taurine (2-amino-ethanesulfonic acid) is one of the most abundant amino acids in the human body. The biosynthesis of taurine is believed to be incomplete in astrocytes and neurons, but metabolic cooperation between these two cell types is essential for the completion of its metabolic pathway.¹ Taurine is ubiquitously distributed, but is enriched in electrically excitable tissues such as the brain, retina, heart, and skeletal muscles.² The regulatory role of taurine has been implicated in a plethora of functions such as an anti-inflammatory molecule,^{3,4} osmolyte, anti-oxidant,^{2,5,6} trophic factor,^{7,8} and as a neuromodulator.⁹⁻¹¹ Clinically, taurine has been used with varying degrees of success for the treatment of a variety of conditions, including, but not limited to, cardiovascular diseases, hypercholesterolemia, epilepsy, macular degeneration, Alzheimer's disease, hepatic disorders, alcoholism, cystic fibrosis, and, most recently in *in vitro* fertilization.^{12,13}

Although taurine is not fully recognized as a neurotransmitter, it satisfies most of the criteria necessary to be classified as one. Co-localization of the taurine-synthesizing enzyme, cysteine sulfinic acid decarboxylase (CSAD), and taurine on the presynaptic side of a nerve has been documented, particularly in association with synaptic vesicles.¹⁴⁻¹⁶ Interestingly, taurine is the only free amino acid that is highly enriched in the synaptic vesicles in comparison with glutamic acid, glutamine, gamma-amino butyric acid (GABA), and aspartic acid, which are also available in the synaptic vesicle fractions.¹⁷ Taurine release is attributed to depolarization-evoked, calcium-dependent pathways and sodium-dependent, calcium-independent pathways under very high potassium concentrations.^{18,19} About two decades ago, we demonstrated the presence of unique taurine receptors, and additional studies have reported the kinetic properties of highly specific taurine receptors that are neither agonists nor antagonists of structurally related amino acids such as glutamate, GABA, or glycine-activated taurine receptors.²⁰ Recently, an independent report further strengthened our findings that a specific recognition site exists that is used exclusively by taurine.²¹ Taurine is known for its ability to neuromodularly inhibit postsynaptic taurine receptors and act as an indirect agonist of GABA_A and glycine receptors, thereby increasing the duration of chloride channel conductance.²²

* Corresponding author. Jang-Yen Wu, Schmidt Senior Fellow and Distinguished Professor, Florida Atlantic University, Charles E. Schmidt College of Medicine, 777 Glades Road, P.O. Box 3091, Boca Raton, FL 33431-0991, USA.
E-mail: J.-Y. Wu <jwu@fau.edu>

Apart from this, the presence of a sodium-dependent taurine transporter (TauT) has been confirmed, and TauT knockouts demonstrate retinal degeneration, reduced olfactory sensitivity, and the manifestation of clinically important age-dependent diseases.^{23–25} It is widely accepted that a biochemical mechanism is required to clear a neurotransmitter from the synaptic cleft after neurotransmission in order to maintain levels below toxicity. For taurine, plasma membrane transporters that are involved in the uptake of taurine from different brain regions, such as the cerebellar regions, the hypothalamus, and neuroglia, have been reported.^{26–28} Although taurine meets the above mentioned criteria, it has been suggested that the presence of a vesicular taurine transporter and the process of vesicular membrane uptake of newly synthesized taurine to be loaded into taurinergic synaptic vesicles clearly defines taurine as a neurotransmitter and, hence, the acceptance of the theory of a taurinergic phenotype. No such evidence supporting the presence of a vesicular taurine transporter or the vesicular uptake of taurine has been documented. In fact, it has been confirmed that aspartate, taurine, and proline are not taken up by any synaptic vesicle, unlike similar amino acids such as glutamate, GABA, and glycine.²⁹ More specific studies are required to examine the proposed putative role of taurine in the central nervous system (CNS). So far, we have examined pertinent information related to the ways in which taurine exerts its neuroprotective effects, and these findings are presented in this review.

Both regenerative medicine and tissue engineering have great potential in clinical medicine because they can completely replace damaged tissue and promote the proliferation and differentiation of terminal cells that cannot otherwise be revived. The cells of the nervous system were once thought to be incapable of regeneration. However, with the success of therapeutic strategies involving the intervention of potent growth factors or cytokines, new cells can be propagated from progenitor cells. For neurons, growth factors such as granulocyte colony-stimulating factor (G-CSF), stromal cell-derived factor-1 (SDF-1), brain-derived neurotrophic factor (BDNF), and glial-derived neurotrophic factor (GDNF) have become increasingly popular for the treatment of a wide spectrum of neurological diseases, including Parkinson's disease, Huntington's disease, neuropathic pain, stroke, etc.^{30–32} Newer reports suggest that G-CSF plays a role in memory impairment in senescence-accelerated mice.³³ G-CSF has been approved by the Food and Drug Administration for clinical use in patients with neutropenia and cancer patients receiving bone marrow transplant, in addition to being used as a novel drug for treating stroke patients.³⁴ G-CSF and its receptors are widely expressed in the neurons of the CNS and, more importantly, in adult neural stem cells.³⁵ Interestingly, G-CSF is able to steadily pass through the blood-brain barrier in intact rats, as demonstrated by a study that utilized G-CSF-iodine dye.³⁶ G-CSF protects against a number of neurological diseases, such as Parkinson's disease,³² Huntington's disease,³⁷ and cerebral ischemia.³⁸ G-CSF stimulates the neural progenitor response *in vivo* and markedly improves long-term behavioral outcomes after cortical ischemia.³¹ Peripheral infusion of G-CSF enhances the recruitment of progenitor cells from the lateral ventricle wall into ischemic areas of the neocortex in rats.³¹ In this review, the molecular mechanisms by which G-CSF contributes to neuroprotection will be discussed.

2. Mechanisms of action of taurine in neuroprotection

2.1. Neuromodulatory role of taurine in the maintenance of intracellular calcium homeostasis

Although normal calcium signaling is crucial for normal physiological functions, calcium dyshomeostasis is a major event in the

pathophysiology of a plethora of neurological diseases, including Alzheimer's disease, cerebral ischemia, Huntington's disease, etc.^{39–41} Cells are endowed with calcium-permeable membrane receptors and channels that are voltage- or ligand-gated, or lodged with ATP-driven pumps such as Na⁺/Ca²⁺ exchangers and plasma membrane Ca²⁺ ATPase, which collectively maintain low levels of intracellular calcium. Excessive activation of glutamate receptors is known to cause a heavy influx and accumulation of calcium inside the cell and is considered as one of the routes that ultimately results in neuronal death. This is mediated by excessive glutamate release because failure to re-uptake calcium by neurons and astroglia has been linked to CNS insults such as traumatic brain injury and Parkinson's disease, to name a couple.⁴² We and other researchers have shown that taurine exerts its protective effects on neurons by effectively regulating intracellular calcium levels. It was initially shown that taurine protects against glutamate-induced neuronal damage by inhibiting the reverse mode of Na⁺-Ca²⁺ exchangers.^{9–11} Further studies have indicated that the protective effects of taurine are also facilitated through L-, P/Q-, and N-type voltage-gated calcium channels and N-Methyl-D-aspartic acid (NMDA) receptors.⁴³ Taurine is also implicated in the inhibition of glutamate-induced release of calcium from internal pools.⁴⁴

2.2. Prevention of glutamate-induced apoptosis by taurine

Glutamatergic neurotransmission is at center stage in neuronal development, differentiation, migration, survival, learning, and memory formation.^{45,46} However, a high concentration of glutamate is associated with the clinical characteristics of various diseases, including stroke, brain trauma, Parkinson's disease, etc.^{42,47} We have reported that taurine prevents glutamate-induced activation of calpain and caspase-9 in rat primary neuronal cultures.⁴⁸ In addition, pre-incubation with taurine prior to glutamate treatment markedly reduced the number of apoptotic cells, as indicated by Hoechst staining, lowered the Bax/Bcl-2 ratio, and attenuated intracellular Ca²⁺ levels.^{9,10,48} A gerbil model of transient focal cerebral ischemia designed to detect alterations in amino acids revealed significant elevations of GABA and taurine, perhaps to combat the surge in posts ischemic glutamate.^{49–51}

2.3. Taurine downregulates key players in the intrinsic apoptosis pathway

Taurine is a strong modulator of apoptosis and is widely known to prevent elevated levels of caspases, calpains, and pro-apoptotic proteins such as Bad, Bax, and Bim. Taurine has been reported to significantly reduce apoptotic death by downregulating the activities of caspase-3 and intracellular calcium.^{52,53} Taurine also represses ischemia-induced caspase-8 and caspase-9 expression in mouse hypothalamic nuclei.⁵⁴ Not only does taurine exercise its anti-apoptotic effects by inhibiting the activation of caspases, but it has also been shown to synergistically upregulate calpastatin while downregulating calpain in a model of focal cerebral ischemia.⁵⁵ A taurine-conjugated form of tauroursodeoxycholic acid (TUDCA) has been shown to be more beneficial than taurine for cell protection. TUDCA reduces the apoptotic threshold induced by glutamate in rat cortical neurons by causing phosphorylation and translocation of Bad from the mitochondria to the cytosol, which is the primary step in inactivating the release of cytochrome c from the mitochondria and triggering the activation of the caspase cascade. TUDCA also appears to modulate, in part, the activation of the PI3 K-dependent Bad signaling pathway.⁵⁶ This also appears to be true in an Alzheimer's disease model of amyloid-beta-induced pathogenesis.⁵⁷ TUDCA has been successfully applied to combat apoptosis-induced Parkinson's and Huntington's disease models.^{58,59}

The cytoprotective role of taurine has been extended to preserving the integrity of mitochondrial pore permeability. Mitochondrial dysfunctions have deleterious consequences on neurons via the increased production of reactive oxygen species (ROS), ATP depletion, and the activation of cell death processes. Based on the current literature, it is apparent that taurine protects against hypoxia-induced apoptosis by preventing mitochondrial dysfunction.⁶⁰ Calcium overload and ionic imbalances in neurons induce mitochondria to produce free radicals.^{61,62} Elevated levels of ROS is a hallmark of neurodegenerative diseases, especially Parkinson's disease.⁶³ Both taurine and TUDCA have been implicated in the significant inhibition of mitochondrial membrane alterations and antagonizing glutamate- and chemical hypoxia-induced calcium overload.^{64–67} Direct evidence supporting taurine's ability to block mitochondrion-mediated cell pathways has been published.⁶⁸ Disruption of the mitochondrial respiration chain leads to cellular swelling followed by osmolyte efflux, as shown by the pathology of stroke.^{69–71} Taurine is known to enhance cell volume regulation when neurons are swollen under extreme pathological conditions.⁷² It has also been shown that taurine efficiently reduces cellular swelling following exposure to oxygen-glucose deprivation and reoxygenation-induced damage in rat brain cortical slices.⁷³

When misfolded or unfolded proteins queue up in the endoplasmic reticulum (ER), the unfolded protein response (UPR) is generated, which then stalls protein synthesis until the proper fold-enhancing molecules are gathered. If the cell is unable to take the quanta of mis- or unfolded proteins, then UPR triggers the caspase-12-mediated apoptotic pathway, which operates exclusively in the ER.⁷⁴ UPR is mediated by ER transmembrane receptor-activating transcription factor 6 (ATF6), inositol-requiring kinase 1 (IRE1),

and double-stranded RNA-activated protein kinase 1 (PKR)-like endoplasmic reticulum kinase (PERK). ER stress is known to be manifested in a variety of brain diseases like Alzheimer's disease, Huntington's chorea, Parkinson's disease, and amyotrophic lateral sclerosis.^{75,76} It is reasonable to believe that cross-talk exists between mitochondria and the ER via the caspase cascade and aberrant calcium signaling. In fact, the synergistic actions of mitochondrial dysfunction and ER stress are both responsible for the pathophysiology of variety of diseases.^{77,78} Our recently published data indicate that taurine protects against glutamate-induced excitotoxicity in primary cortical neurons and hypoxia-induced toxicity in PC12 cells by downregulating the expression of CHOP, GRP78, Bim, and caspase-12, which are the key proteins related to ER stress.^{79,80}

2.4. Taurine counteracts excitotoxic upsurges by interacting with GABAA and glycine receptors, thereby increasing the duration of chloride conductance

Several lines of evidence indicate that taurine inhibits neurotransmission by binding to ionotropic GABAA and glycine receptors; this has been effectively used for the treatment of Alzheimer's disease.^{22,81} Taurine conducts the flow of chloride not by increasing the frequency of the opening of the chloride channels, but by increasing the duration that the channel is open.⁸² Very few studies have been conducted on the direct activation of taurine receptors. Definitive identification of taurine receptors is still emerging.^{20,21} Further mechanistic studies are necessary to understand the direct role of taurine receptors on propagating neurotransmission. A summary schematic depicting the mode of action of taurine as a neuroprotective agent is shown in Figure 1.

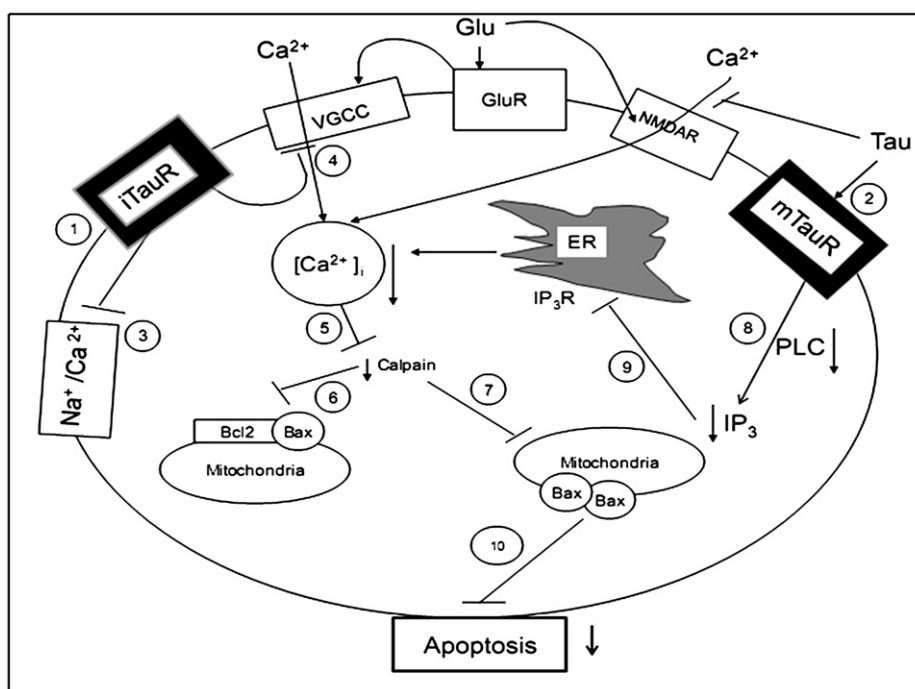


Figure 1 Summary schematic depicting the mode of action of taurine in neuroprotection. The sequence of events leading from the activation of taurine receptors to neuroprotection can be summarized as follows: (1) activation of ionotropic taurine receptors (iTauR) and/or activation of metabotropic taurine receptors (mTauR); (2) inhibition of the reverse mode of sodium/calcium exchangers; (3) inhibition of voltage-gated calcium channels (VGCC) by taurine-induced hyperpolarization; (4) inhibition of calpain resulting from the decrease in the intracellular free-calcium concentration; (5) inhibition of the cleavage of Bcl-2 and Bax by the inhibition of calpain; (6) inhibition of the formation of the Bax homodimer, leading to the inhibition of apoptosis; (7) activation of mTauR, which is negatively coupled to inhibitory G proteins, resulting in the inhibition of phospholipase C (PLC) activity and a decrease in IP₃ production; (8) decreased IP₃ level inhibits the release of calcium from the internal calcium storage pools, such as the ER, resulting in reduced ER stress and inhibition of apoptosis.

3. Mode of action of G-CSF in neuroprotection

G-CSF is a growth factor that is known to stimulate the proliferation and survival of hematopoietic cells.⁸³ G-CSF can penetrate the blood-brain barrier and plays a prominent role in the CNS.³⁵ G-CSF and its receptors are expressed in neurons throughout the brain and their expression is induced by ischemia, which is suggestive of an autocrine protective signaling mechanism.³⁵ An increasing amount of evidence indicates that G-CSF is neuroprotective and neuroregenerative both *in vivo* and *in vitro*. For example, G-CSF protects against neurodegeneration in a number of neurological disease models, such as Parkinson's disease,^{32,84,85} Huntington's disease,³⁷ and cerebral ischemia.⁸⁶ The neuroprotective functions of G-CSF are further discussed below.

3.1. Suppression of multiple apoptotic pathways

G-CSF has been consistently cited as an attenuator of apoptosis. G-CSF is known to reduce the number of apoptotic cells identified by cleaved caspase-3-immunoreactive neurons and terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL)-positive cells in neonatal hypoxic-ischemic rats.⁸⁷ G-CSF also extends its protective effects by downregulating a number of anti-apoptotic factors such as Bax, caspase-3, upregulated Bcl-2, Bcl-xL, and Pim-1, thereby synergistically preventing the release of cytochrome into the cytosol and translocating Bax to the mitochondria,^{88–91} as shown in Figure 2.

In addition, G-CSF does not only modulate the intrinsic apoptosis pathway, but also the extrinsic apoptosis pathway by mediating its anti-apoptotic role through the tumor necrosis factor-related, apoptosis-inducing ligand (TRAIL) pathway.³³ Recombinant G-CSF

reduces the number of TRAIL-positive neurons and protects senescence-accelerated mice against memory impairment.³³ Such properties make G-CSF an attractive therapy for the treatment of diseases characterized by dementia. In retinal ganglion cells, the anti-apoptotic properties of G-CSF have been attributed to the Phosphatidylinositol 3-kinase (PI3)/AKT pathway and 6-hydroxydopamine (6-OHDA)-induced toxicity via the ERK pathway.^{92,93} Recently, we reported that G-CSF alone, or in combination with taurine, protects glutamate-induced primary rat neuronal cultures by downregulating the ER stress markers GRP78, CHOP, Bim, and caspase-12 *in vitro*.⁸⁰ In addition to its neuroprotective functions, G-CSF also exerts effects on the neuroregenerative/stem cell mechanisms, as discussed in the following section.

3.2. Activation of cell proliferation mechanisms that promote neurogenesis

G-CSF stimulates the neural progenitor response *in vivo* and markedly improves long-term behavioral outcomes after cortical ischemia.³⁵ Peripheral infusion of G-CSF enhances the recruitment of progenitor cells from the lateral ventricle wall into ischemic areas of the neocortex in rats.³⁵ Furthermore, G-CSF is known to induce neurogenesis by activating signal transducer and activator of transcription 3 (STAT3), signal transducer and activator of transcription 5 (STAT5), and vascular endothelial growth factor (VEGF).^{87,94,95} In an *in vivo* 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) model of Parkinson's disease, G-CSF significantly increases the number of dopamine (DA) neurons and the functions of the DA system, suggesting that G-CSF restores the degenerated nervous system through both neuroprotective and neurogenetic mechanisms.⁸⁴

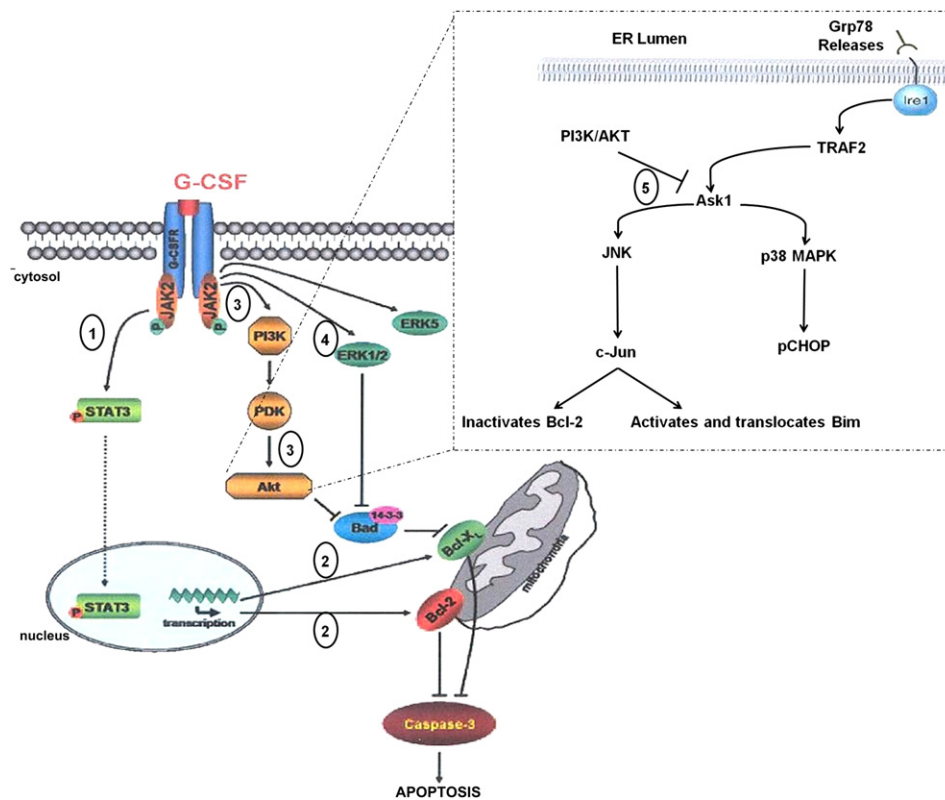


Figure 2 Proposed mode of action of the neuroprotective functions of G-CSF. G-CSF could exert its neuroprotective functions through one or more of the following signaling pathways: (1) activation of the STAT3 pathway results in translocation of STAT3 to the nucleus and (2) the upregulation of the anti-apoptotic genes, *Bcl-2* and *Bcl-X*; (3) activation of the PI3 K/AKT pathways or (4) ERK1/2 pathway results in the inhibition of the pro-apoptotic protein, Bad; (5) activation of the PI3 K/AKT pathways inhibits the ER stress-mediated ASK-1 pathway, resulting in disinhibition or activation of the anti-apoptotic protein, Bcl-2, and inactivation of the pro-apoptotic protein, BIM (see insert).

3.3. Clinical applications of G-CSF

In our recent studies, we have observed a remarkable improvement in neuronal functions, both in clinical cases and Parkinson's disease animal models that have been treated with G-CSF. Among the clinical cases, a patient with end-stage corticobasal ganglionic degeneration showed marked improvement after G-CSF treatment based on the patient's evaluation according to the Unified Parkinson's Disease Rating System (UPDRS). Overall improvement was 66% across all four categories: 1) mentation, behavior, and mood; 2) daily life activities; 3) motor skills; and 4) complications.⁸⁵ These results suggest that G-CSF may promote the regeneration of DA neurons in the substantia nigra and their functional integration into the nigrostriatal pathway. To extend this work, we conducted laboratory tests using the MPTP mouse model of Parkinson's disease. Unlike other published studies where G-CSF was administered before MPTP treatment,³² in our study we opted for delayed treatment with G-CSF until after degeneration of the DA neurons by MPTP had been completed. We found that MPTP causes a marked loss in DA neurons, and G-CSF treatment restores the functions of the DA system, as indicated by increases in the number of DA neurons, stimulation-induced DA release, restoration of the nigrostriatal pathway, and improvement in locomotor activities, all of which are suggestive that the observed restoration might be due to differentiation of substantia nigra neuronal progenitor cells or progenitor cells that invade the substantia nigra after G-CSF application.^{84,85} In addition to PD, we also found that G-CSF markedly reduces the size of brain infarctions that are induced by middle cerebral artery occlusion stroke animal model. These findings support the notion that G-CSF is a novel agent with both neuroregenerative/stem cell and neuroprotective activities and could be effective for the treatment of Parkinson's disease, stroke, and other degenerative brain disorders.

4. Conclusion

In the present review, we demonstrate that both taurine and G-CSF are attractive therapeutic targets for the treatment of neurodegenerative diseases. Both taurine and G-CSF function by suppressing apoptosis at multiple levels. The synergistic action of the combination of taurine and G-CSF has been proven to be beneficial for treating glutamate-induced neurotoxicity in primary rat neuronal cultures.⁸⁰ Because taurine is a natural amino acid and G-CSF is approved by the Food and Drug Administration, these findings could push forward the development of combinational approaches that provide more effective therapies.

Acknowledgments

This work was supported, in part, by the James and Esther King Biomedical Research Program, Florida Department of Health (grant #: 09KW-11), and the Schmidt Foundation, Charles E. Schmidt College of Medicine, Florida Atlantic University.

References

- Vitvitsky V, Garg SK, Banerjee R. Taurine biosynthesis by neurons and astrocytes. *J Biol Chem* 2011;**286**:32002–10.
- Oja SS, Saransaari P. Pharmacology of taurine. *Proc West Pharmacol* 2007;**50**:8–15.
- Sun M, Zhao Y, Gu Y, Xu C. Anti-inflammatory mechanism of taurine against ischemic stroke is related to down-regulation of PARP and NF- κ B. *Amino Acids* 2011.
- Miao J, Zhang J, Zheng L, Yu X, Zhu W, Zou S. Taurine attenuates Streptococcus uberis-induced mastitis in rats by increasing T regulatory cells. *Amino Acids* 2011.
- Schaffer S, Takahashi K, Azuma J. Role of osmoregulation in the actions of taurine. *Amino Acids* 2000;**19**:527–46.
- Schaffer SW, Azuma J, Mozaffari M. Role of antioxidant activity of taurine in diabetes. *Can J Physiol Pharmacol* 2009;**87**:91–9.
- Hernandez-Benitez R, Pasantes-Morales H, Saldana IT, Ramos-Mandujano G. Taurine stimulates proliferation of mice embryonic cultured neural progenitor cells. *J Neurosci Res* 2010;**88**:1673–81.
- Lima L, Cubillos S. Taurine might be acting as a trophic factor in the retina by modulating phosphorylation of cellular proteins. *J Neurosci Res* 1998;**53**:377–84.
- Chen WQ, Jin H, Nguyen M, Carr J, Lee YJ, Hsu CC, Faiman MD, et al. Role of taurine in regulation of intracellular calcium level and neuroprotective function in cultured neurons. *J Neurosci Res* 2001;**66**:612–9.
- El Idrissi A, Trenkner E. Growth factors and taurine protect against excitotoxicity by stabilizing calcium homeostasis and energy metabolism. *J Neurosci* 1999;**19**:9459–68.
- Foos TM, Wu JY. The role of taurine in the central nervous system and the modulation of intracellular calcium homeostasis. *Neurochem Res* 2002;**27**:21–6.
- Bidri M, Choay P. Taurine: a particular aminoacid with multiple functions. *Ann Pharm Fr* 2003;**61**:385–91.
- Birdsall TC. Therapeutic applications of taurine. *Altern Med Rev* 1998;**3**:128–36.
- Lin CT, Song GX, Wu JY. Ultrastructural demonstration of L-glutamate decarboxylase and cysteinesulfinic acid decarboxylase in rat retina by immunocytochemistry. *Brain Res* 1985;**331**:71–80.
- Magnusson KR, Clements JR, Wu JY, Beitz AJ. Colocalization of taurine- and cysteine sulfinic acid decarboxylase-like immunoreactivity in the hippocampus of the rat. *Synapse* 1989;**4**:55–69.
- Magnusson KR, Madl JE, Clements JR, Wu JY, Larson AA, Beitz AJ. Colocalization of taurine- and cysteine sulfinic acid decarboxylase-like immunoreactivity in the cerebellum of the rat with monoclonal antibodies against taurine. *J Neurosci* 1988;**8**:4551–64.
- Kontro P, Marnela KM, Oja SS. Free amino acids in the synaptosome and synaptic vesicle fractions of different bovine brain areas. *Brain Res* 1980;**184**:129–41.
- Philibert RA, Rogers KL, Dutton GR. Stimulus-coupled taurine efflux from cerebellar neuronal cultures: on the roles of Ca⁺⁺ and Na⁺. *J Neurosci Res* 1989;**22**:167–71.
- Pin JP, Weiss S, Sebben M, Kemp DE, Bockaert J. Release of endogenous amino acids from striatal neurons in primary culture. *J Neurochem* 1986;**47**:594–603.
- Wu JY, Tang XW, Tsai WH. Taurine receptor: kinetic analysis and pharmacological studies. *Adv Exp Med Biol* 1992;**315**:263–8.
- Frosini M, Sesti C, Saponara S, Ricci L, Valoti M, Palmi M, Machetti F, et al. A specific taurine recognition site in the rabbit brain is responsible for taurine effects on thermoregulation. *Br J Pharmacol* 2003;**139**:487–94.
- Wu J, Kohno T, Georgiev SK, Ikoma M, Ishii H, Petrenko AB, Baba H. Taurine activates glycine and gamma-aminobutyric acid A receptors in rat substantia gelatinosa neurons. *Neuroreport* 2008;**19**:333–7.
- Heller-Stilb B, van Roeyen C, Rascher K, Hartwig HG, Huth A, Seeliger MW, Warskulat U, et al. Disruption of the taurine transporter gene (taut) leads to retinal degeneration in mice. *FASEB J* 2002;**16**:231–3.
- Warskulat U, Borsch E, Reinehr R, Heller-Stilb B, Roth C, Witt M, Haussinger D. Taurine deficiency and apoptosis: findings from the taurine transporter knockout mouse. *Arch Biochem Biophys* 2007;**462**:202–9.
- Warskulat U, Heller-Stilb B, Oermann E, Zilles K, Haas H, Lang F, Haussinger D. Phenotype of the taurine transporter knockout mouse. *Methods Enzymol* 2007;**428**:439–58.
- Besson MT, Re DB, Moulin M, Birman S. High affinity transport of taurine by the Drosophila aspartate transporter dEAAT2. *J Biol Chem* 2005;**280**:6621–6.
- Chan-Palay V, Lin CT, Palay S, Yamamoto M, Wu JY. Taurine in the mammalian cerebellum: demonstration by autoradiography with [³H]taurine and immunocytochemistry with antibodies against the taurine-synthesizing enzyme, cysteine-sulfinic acid decarboxylase. *Proc Natl Acad Sci (USA)* 1982;**79**:2695–9.
- Hanretta AT, Lombardini JB. Is taurine a hypothalamic neurotransmitter? A model of the differential uptake and compartmentalization of taurine by neuronal and glial cell particles from the rat hypothalamus. *Brain Res* 1987;**434**:167–201.
- Fykse EM, Fonnum F. Amino acid neurotransmission: dynamics of vesicular uptake. *Neurochem Res* 1996;**21**:1053–60.
- Hess DC, Borlongan CV. Stem cells and neurological diseases. *Cell Prolif* 2008;**41**(suppl. 1):94–114.
- Schneider A, Kuhn HG, Schabitz WR. A role for G-CSF (granulocyte-colony stimulating factor) in the central nervous system. *Cell Cycle* 2005;**4**:1753–7.
- Meuer K, Pitzer C, Teismann P, Kruger C, Goricke B, Laage R, Lingor P, et al. Granulocyte-colony stimulating factor is neuroprotective in a model of Parkinson's disease. *J Neurochem* 2006;**97**:675–86.
- Zhao C, Xie Z, Wang P, Wang Y, Lai C, Zhu Z, Liu Z, et al. Granulocyte-colony stimulating factor protects memory impairment in the senescence-accelerated mouse (SAM)-P10. *Neuro Res* 2011;**33**:354–9.
- Bennett CL, Smith TJ, Weeks JC, Bredt AB, Feinglass J, Fetting JH, Hillner BE, et al. Use of hematopoietic colony-stimulating factors: the American Society of Clinical Oncology survey. The Health Services Research Committee of the American Society of Clinical Oncology. *J Clin Oncol* 1996;**14**:2511–20.

35. Schneider A, Kruger C, Steigleder T, Weber D, Pitzer C, Laage R, Aronowski J, et al. The hematopoietic factor G-CSF is a neuronal ligand that counteracts programmed cell death and drives neurogenesis. *J Clin Invest* 2005;**115**:2083–98.
36. Zhao LR, Navalitloha Y, Singhal S, Mehta J, Piao CS, Guo WP, Kessler JA, et al. Hematopoietic growth factors pass through the blood-brain barrier in intact rats. *Exp Neurol* 2007;**204**:569–73.
37. Lee ST, Park JE, Kim DH, Kim S, Im WS, Kang L, Jung SH, et al. Granulocyte-colony stimulating factor attenuates striatal degeneration with activating survival pathways in 3-nitropropionic acid model of Huntington's disease. *Brain Res* 2008;**1194**:130–7.
38. Sevimli S, Diederich K, Strecker JK, Schilling M, Klocke R, Nikol S, Kirsch F, et al. Endogenous brain protection by granulocyte-colony stimulating factor after ischemic stroke. *Exp Neurol* 2009;**217**:328–35.
39. Paschen W. Dependence of vital cell function on endoplasmic reticulum calcium levels: implications for the mechanisms underlying neuronal cell injury in different pathological states. *Cell Calcium* 2001;**29**:1–11.
40. Pivovarova NB, Andrews SB. Calcium-dependent mitochondrial function and dysfunction in neurons. *FEBS J* 2010;**277**:3622–36.
41. Tang TS, Slow E, Lupu V, Stavrovskaya IG, Sugimori M, Llinas R, Kristal BS, et al. Disturbed Ca²⁺ signaling and apoptosis of medium spiny neurons in Huntington's disease. *Proc Natl Acad Sci U S A* 2005;**102**:2602–7.
42. Lau A, Tymianski M. Glutamate receptors, neurotoxicity and neurodegeneration. *Pflugers Arch* 2010;**460**:525–42.
43. Wu H, Jin Y, Wei J, Jin H, Sha D, Wu JY. Mode of action of taurine as a neuro-protector. *Brain Res* 2005;**1038**:123–31.
44. Wu JY, Chen W, Tang XW, Jin H, Foss T, Schloss JV, Davis K, et al. Mode of action of taurine and regulation dynamics of its synthesis in the CNS. *Adv Exp Med Biol* 2000;**483**:35–44.
45. Behar TN, Scott CA, Greene CL, Wen X, Smith SV, Maric D, Liu QY, et al. Glutamate acting at NMDA receptors stimulates embryonic cortical neuronal migration. *J Neurosci* 1999;**19**:4449–61.
46. Riedel G, Platt B, Micheau J. Glutamate receptor function in learning and memory. *Behav Brain Res* 2003;**140**:1–47.
47. Wang Y, Qin ZH. Molecular and cellular mechanisms of excitotoxic neuronal death. *Apoptosis* 2010;**15**:1382–402.
48. Leon R, Wu H, Jin Y, Wei J, Buddhala C, Prentice H, Wu JY. Protective function of taurine in glutamate-induced apoptosis in cultured neurons. *J Neurosci Res* 2009;**87**:1185–94.
49. Khan SH, Banigesh A, Baziani A, Todd KG, Miyashita H, Eweida M, Shuaib A. The role of taurine in neuronal protection following transient global forebrain ischemia. *Neurochem Res* 2000;**25**:217–23.
50. Tang XC, Rao MR, Hu G, Wang H. Alterations of amino acid levels from striatum, hippocampus, and cerebral cortex induced by global cerebral ischemia in gerbil. *Acta Pharmacol Sin* 2000;**21**:819–23.
51. Saransaari P, Oja SS. Modulation of taurine release in ischemia by glutamate receptors in mouse brain stem slices. *Amino Acids* 2010;**38**:739–46.
52. Gao X, Yang X, Zhang B. Neuroprotection of taurine against bilirubin-induced elevation of apoptosis and intracellular free calcium ion. *in vivo. Toxicol Mech Methods* 2011;**21**:383–7.
53. Taranukhin AG, Taranukhina EY, Saransaari P, Podkletnova IM, Pelto-Huikko M, Oja SS. Neuroprotection by taurine in ethanol-induced apoptosis in the developing cerebellum. *J Biomed Sci* 2010;**17**(suppl. 1):S12.
54. Taranukhin AG, Taranukhina EY, Saransaari P, Djatchkova IM, Pelto-Huikko M, Oja SS. Taurine reduces caspase-8 and caspase-9 expression induced by ischemia in the mouse hypothalamic nuclei. *Amino Acids* 2008;**34**:169–74.
55. Sun M, Xu C. Neuroprotective mechanism of taurine due to up-regulating calpastatin and down-regulating calpain and caspase-3 during focal cerebral ischemia. *Cell Mol Neurobiol* 2008;**28**:593–611.
56. Castro RE, Sola S, Ramalho RM, Steer CJ, Rodrigues CM. The bile acid tauroursodeoxycholic acid modulates phosphorylation and translocation of bad via phosphatidylinositol 3-kinase in glutamate-induced apoptosis of rat cortical neurons. *J Pharmacol Exp Ther* 2004;**311**:845–52.
57. Sola S, Castro RE, Laires PA, Steer CJ, Rodrigues CM. Tauroursodeoxycholic acid prevents amyloid-beta peptide-induced neuronal death via a phosphatidylinositol 3-kinase-dependent signaling pathway. *Mol Med* 2003;**9**:226–34.
58. Duan WM, Rodrigues CM, Zhao LR, Steer CJ, Low WC. Tauroursodeoxycholic acid improves the survival and function of nigral transplants in a rat model of Parkinson's disease. *Cell Transplant* 2002;**11**:195–205.
59. Keene CD, Rodrigues CM, Eich T, Linehan-Stieers C, Abt A, Kren BT, Steer CJ, et al. A bile acid protects against motor and cognitive deficits and reduces striatal degeneration in the 3-nitropropionic acid model of Huntington's disease. *Exp Neurol* 2001;**171**:351–60.
60. Chen K, Zhang Q, Wang J, Liu F, Mi M, Xu H, Chen F, et al. Taurine protects transformed rat retinal ganglion cells from hypoxia-induced apoptosis by preventing mitochondrial dysfunction. *Brain Res* 2009;**1279**:131–8.
61. Dykens JA. Isolated cerebral and cerebellar mitochondria produce free radicals when exposed to elevated CA²⁺ and Na⁺: implications for neurodegeneration. *J Neurochem* 1994;**63**:584–91.
62. Lievre V, Becuwe P, Bianchi A, Bossenmeyer-Pourie C, Koziel V, Franck P, Nicolas MB, et al. Intracellular generation of free radicals and modifications of detoxifying enzymes in cultured neurons from the developing rat forebrain in response to transient hypoxia. *Neuroscience* 2001;**105**:287–97.
63. Jana S, Sinha M, Chanda D, Roy T, Banerjee K, Munshi S, Patro BS, et al. Mitochondrial dysfunction mediated by quinone oxidation products of dopamine: implications in dopamine cytotoxicity and pathogenesis of Parkinson's disease. *Biochim Biophys Acta* 2011;**1812**:663–73.
64. Rodrigues CM, Stieers CL, Keene CD, Ma X, Kren BT, Low WC, Steer CJ. Tauroursodeoxycholic acid partially prevents apoptosis induced by 3-nitropropionic acid: evidence for a mitochondrial pathway independent of the permeability transition. *J Neurochem* 2000;**75**:2368–79.
65. Zhao P, Huang YL, Cheng JS. Taurine antagonizes calcium overload induced by glutamate or chemical hypoxia in cultured rat hippocampal neurons. *Neurosci Lett* 1999;**268**:25–8.
66. El Idrissi A. Taurine increases mitochondrial buffering of calcium: role in neuroprotection. *Amino Acids* 2008;**34**:321–8.
67. Palmi M, Youmbi GT, Sgaragli G, Meini A, Benocci A, Fusi F, Frosini M, et al. The mitochondrial permeability transition and taurine. *Adv Exp Med Biol* 2000;**483**:87–96.
68. Sun M, Gu Y, Zhao Y, Xu C. Protective functions of taurine against experimental stroke through depressing mitochondria-mediated cell death in rats. *Amino Acids* 2011;**40**:1419–29.
69. Patel AJ, Lauritzen I, Lazdunski M, Honore E. Disruption of mitochondrial respiration inhibits volume-regulated anion channels and provokes neuronal cell swelling. *J Neurosci* 1998;**18**:3117–23.
70. Harvey VL, Saul MW, Garner C, McDonald RL. A role for the volume regulated anion channel in volume regulation in the murine CNS cell line, CAD. *Acta Physiol (Oxf)* 2010;**198**:159–68.
71. Pasantes-Morales H, Tuz K. Volume changes in neurons: hyperexcitability and neuronal death. *Contrib Nephrol* 2006;**152**:221–40.
72. Kreisman NR, Olson JE. Taurine enhances volume regulation in hippocampal slices swollen osmotically. *Neuroscience* 2003;**120**:635–42.
73. Ricci L, Valoti M, Sgaragli G, Frosini M. Protection by taurine of rat brain cortical slices against oxygen glucose deprivation- and reoxygenation-induced damage. *Eur J Pharmacol* 2009;**621**:26–32.
74. Chakrabarti A, Chen AW, Varner JD. A review of the mammalian unfolded protein response. *Biotechnol Bioeng* 2011;**108**:2777–93.
75. Paschen W, Mengesdorf T. Endoplasmic reticulum stress response and neurodegeneration. *Cell Calcium* 2005;**38**:409–15.
76. Martinez JA, Zhang Z, Svetlov SI, Hayes RL, Wang KK, Larner SF. Calpain and caspase processing of caspase-12 contribute to the ER stress-induced cell death pathway in differentiated PC12 cells. *Apoptosis* 2010;**15**:1480–93.
77. Ilieva EV, Kichev A, Naudi A, Ferrer I, Pamplona R, Portero-Otin M. Mitochondrial dysfunction and oxidative and endoplasmic reticulum stress in argyrophilic grain disease. *J Neuropathol Exp Neurol* 2011;**70**:253–63.
78. Smaili S, Hirata H, Ureshino R, Monteforte PT, Morales AP, Muler ML, Terashima J, et al. Calcium and cell death signaling in neurodegeneration and aging. *An Acad Bras Cienc* 2009;**81**:467–75.
79. Pan C, Giraldo GS, Prentice H, Wu JY. Taurine protection of PC12 cells against endoplasmic reticulum stress induced by oxidative stress. *J Biomed Sci* 2010;**17**(suppl 1):S17.
80. Pan C, Gupta A, Prentice H, Wu JY. Protection of taurine and granulocyte colony-stimulating factor against excitotoxicity induced by glutamate in primary cortical neurons. *J Biomed Sci* 2010;**17**(suppl. 1):S18.
81. Louzada PR, Paula Lima AC, Mendonca-Silva DL, Noel F, De Mello FG, Ferreira ST. Taurine prevents the neurotoxicity of beta-amyloid and glutamate receptor agonists: activation of GABA receptors and possible implications for Alzheimer's disease and other neurological disorders. *FASEB J* 2004;**18**:511–8.
82. Okamoto K, Kimura H, Sakai Y. Taurine-induced increase of the Cl⁻ conductance of cerebellar Purkinje cell dendrites in vitro. *Brain Res* 1983;**259**:319–23.
83. Demetri GD, Griffin JD. Granulocyte colony-stimulating factor and its receptor. *Blood* 1991;**78**:2791–808.
84. McCollum M, Ma Z, Cohen E, Leon R, Tao R, Wu JY, Maharaj D, et al. Post-MPTP treatment with granulocyte colony-stimulating factor improves nigrostriatal function in the mouse model of Parkinson's disease. *Mol Neurobiol* 2010;**41**:410–9.
85. Wu JY, Maharaj D. *Method of treating neurodegenerative diseases*. United States Patent and Trademark Office; 2008. Publication No. US-2008-0300176-A1.
86. Lu CZ, Xiao BG. Neuroprotection of G-CSF in cerebral ischemia. *Front Biosci* 2007;**12**:2869–75.
87. Yata K, Matchett GA, Tsubokawa T, Tang J, Kanamaru K, Zhang JH. Granulocyte-colony stimulating factor inhibits apoptotic neuron loss after neonatal hypoxia-ischemia in rats. *Brain Res* 2007;**1145**:227–38.
88. Nishio Y, Koda M, Kamada T, Someya Y, Kadota R, Mannoji C, Miyashita T, et al. Granulocyte colony-stimulating factor attenuates neuronal death and promotes functional recovery after spinal cord injury in mice. *J Neuropathol Exp Neurol* 2007;**66**:724–31.
89. Pitzer C, Klusmann S, Kruger C, Letellier E, Plass C, Dittgen T, Kirsch F, et al. The hematopoietic factor granulocyte-colony stimulating factor improves outcome in experimental spinal cord injury. *J Neurochem* 2010;**113**:930–42.
90. Solaroglu I, Tsubokawa T, Cahill J, Zhang JH. Anti-apoptotic effect of granulocyte-colony stimulating factor after focal cerebral ischemia in the rat. *Neuroscience* 2006;**143**:965–74.
91. Henriques A, Pitzer C, Dupuis L, Schneider A. G-CSF protects motoneurons against axotomy-induced apoptotic death in neonatal mice. *BMC Neurosci* 2010;**11**:25.

92. Tsai RK, Chang CH, Sheu MM, Huang ZL. Anti-apoptotic effects of human granulocyte colony-stimulating factor (G-CSF) on retinal ganglion cells after optic nerve crush are PI3K/AKT-dependent. *Exp Eye Res* 2010;**90**:537–45.
93. Huang HY, Lin SZ, Kuo JS, Chen WF, Wang MJ. G-CSF protects dopaminergic neurons from 6-OHDA-induced toxicity via the ERK pathway. *Neurobiol Aging* 2007;**28**:1258–69.
94. Jung KH, Chu K, Lee ST, Kim SJ, Sinn DI, Kim SU, Kim M, et al. Granulocyte colony-stimulating factor stimulates neurogenesis via vascular endothelial growth factor with STAT activation. *Brain Res* 2006;**1073-1074**:190–201.
95. Schabitz WR, Kollmar R, Schwaninger M, Juettler E, Bardutzky J, Scholzke MN, Sommer C, et al. Neuroprotective effect of granulocyte colony-stimulating factor after focal cerebral ischemia. *Stroke* 2003;**34**:745–51.